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NEWS 19 MAR 16 CASREACT coverage extended
NEWS 20 MAR 20 MARPAT now updated daily
NEWS 21 MAR 22 LWPI reloaded
NEWS 22 MAR 30 RDISCLOSURE reloaded with enhancements
NEWS 23 APR 02 JICST-EPLUS removed from database clusters and STN
NEWS 24 APR 30 GENBANK reloaded and enhanced with Genome Project ID field
NEWS 25 APR 30 CHEMCATS enhanced with 1.2 million new records
NEWS 26 APR 30 CA/CAplus enhanced with 1870-1889 U.S. patent records
NEWS 27 APR 30 INPADOC replaced by INPADOCDB on STN
NEWS 28 MAY 01 New CAS web site launched
NEWS 29 MAY 08 CA/CAplus Indian patent publication number format defined
NEWS 30 MAY 14 RDISCLOSURE on STN Easy enhanced with new search and display fields
NEWS 31 MAY 21 BIOSIS reloaded and enhanced with archival data
NEWS 32 MAY 21 TOXCENTER enhanced with BIOSIS reload
NEWS 33 MAY 21 CA/CAplus enhanced with additional kind codes for German patents
NEWS 34 MAY 22 CA/CAplus enhanced with IPC reclassification in Japanese patents

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AND CURRENT DISCOVER FILE IS DATED 25 SEPTEMBER 2006.

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=> s glutamine synthetase
L1 18104 GLUTAMINE SYNTHETASE

=> s l1 and marker
L2 793 L1 AND MARKER

=> s l2 and CMV
L3 9 L2 AND CMV

=> dup rem l3
PROCESSING COMPLETED FOR L3
L4 9 DUP REM L3 (0 DUPLICATES REMOVED)

=> d bib abs 1-
YOU HAVE REQUESTED DATA FROM 9 ANSWERS - CONTINUE? Y/(N):y

L4 ANSWER 1 OF 9 CAPLUS COPYRIGHT 2007 ACS on STN
AN 2007:175345 CAPLUS <<LOGINID::20070611>>
DN 146:222528
TI Expression vector and methods of producing erythropoietin and other recombinant proteins in mammalian cells for potential therapeutic use
IN Singh, Arun K.; Goel, Ashish; Mendiratta, Sanjeev K.
PA Cadila Healthcare Limited, India
SO PCT Int. Appl., 33pp.
CODEN: PIXXD2
DT Patent
LA English
FAN.CNT 1

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI WO 2007017903	A2	20070215	WO 2006-IN207	20060619

W: AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BW, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, EG, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KM, KN, KP, KR, KZ, LA, LC, LK, LR, LS, LT, LU, LV, LY, MA, MD, MG, MK, MN, MW, MX, MZ, NA, NG, NI, NO, NZ, OM, PG, PH, PL, PT, RO, RS, RU, SC, SD, SE, SG, SK, SL, SM, SY, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, ZA, ZM, ZW
RW: AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HU, IE, IS, IT, LT, LU, LV, MC, NL, PL, PT, RO, SE, SI, SK, TR, BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG, BW, GH, GM, KE, LS, MW, MZ, NA, SD, SL, SZ, TZ, UG, ZM, ZW, AM, AZ, BY, KG, KZ, MD, RU, TJ, TM

PRAI IN 2005-MU720 A 20050620
AB The present invention provides an expression vector and methods for overproducing erythropoietin and other recombinant proteins such as TNFR-IgGFc fusion proteins, rituximab, trastuzumab, bevacuzumab, or other monoclonal antibodies in mammalian cells. The expression vector comprises a ***CMV*** promoter or functional variants, an intron, TPL regulatory element or its functional variants, VA genes or functional variants, and a bovine growth hormone polyadenylation element or functional variants. Upon stable transfection of CHO-DHFR- cells, the expression vector provides 11,830 IU/mL in 168 h culture, which is equiv. to 18.2 to 27.3 .mu.g/106 cells/24 h. This expression vectors provides 80-100% higher expression of recombinant erythropoietin than those previously reported in ther literature.

L4 ANSWER 2 OF 9 BIOSIS COPYRIGHT (c) 2007 The Thomson Corporation on STN
AN 2007:200562 BIOSIS <<LOGINID::20070611>>
DN PREV200700197446
TI Derivation and characterization of cholesterol-independent non-GS NS0 cell lines for production of recombinant antibodies.
AU Hartman, Taymar E. [Reprint Author]; Sar, Nalin; Genereux, Kimberly; Barritt, Diana S.; He, Yimin; Burky, John E.; Wesson, Mark C.; Tso, J. Yun; Tsurushita, Naoya; Zhou, Weichang; Sauer, Paul W.
CS PDL BioPharma Inc, Proc Sci and Engn, 34801 Campus Dr, Fremont, CA 94555
USA
taymar.hartman@pdl.com
SO Biotechnology and Bioengineering, (FEB 1 2007) Vol. 96, No. 2, pp. 294-306.
CODEN: BIBIAU. ISSN: 0006-3592.
DT Article
LA English
ED Entered STN: 21 Mar 2007
Last Updated on STN: 21 Mar 2007
AB Presented is an antibody production platform based on the fed-batch culture of recombinant NSO-derived cell lines. NSO host cells, obtained from the European Collection of Cell Cultures (ECACC, Salisbury, UK, Part No. 85110503), were first adapted to grow in a protein-free; cholesterol-free medium. The resulting host cell line was designated NS0-PFCF (protein-free, cholesterol-free). The five production cell lines presented here were generated using a common protocol consisting of transfection by electroporation and subcloning. The NS0-PFCF host cell line was transfected using a single expression vector contain ing the

Escherichia coli xanthine-guanine phosphoribosyl transferase gene (gpt), and the antibody heavy and light chain genes driven by the ***CMV*** promoter. The five cell lines were chosen after one to three rounds of iterative subclon ing, which resulted in a 19-64% increase in antibody productivity when four mother-daughter cell pairs were cultured in a fed-batch bioreactor process. The production cell lines were genetically characterized to determine antibody gene integrity, nucleotide sequences, copy number, and the number of insertion sites in the NS0 cell genome. Genetic characterization data indicate that each of the five production cell lines, has a single stably integrated copy of the antibody expression vector, and that the antibody genes are correctly expressed. Stability of antibody production was evaluated for three of the five cell lines by comparing the early stage seed bank with the Working Cell Bank (WCB). Antibody productivity was shown to be stable in two of three cell lines evaluated; while one of the cell lines exhibited a 20% drop in productivity after passaging for approximately 4 weeks. These five NSO-derived production cell lines were successfully cultured to produce antibodies with acceptable product quality attributes in a standardized fed-batch bior- eactor process, consistently achieving an average specific productivity of 20-60 pg/cell-day, and a volumetric productivity exceeding 120. mg/L-day (Burky et al., 2006),. In contrast to the commonly available NSO host cell line, which requires serum and cholesterol for growth; and the commonly used expression vector system, which uses a proprietary ***glutamine*** ***synthetase*** selection ***marker*** (GS-NSO), these NSO cells are cholesterol-independent, grow well in a protein-free medium, use a non-proprietary selection ***marker*** , and do not require gene amplification for productivity improvement: These characteristics are advantageous for use of this NSO cell line platform for manufacturing therapeutic antibodies.

L4 ANSWER 3 OF 9 EMBASE COPYRIGHT (c) 2007 Elsevier B.V. All rights reserved on STN
AN 2007081036 EMBASE <<LOGINID::20070611>>
TI Derivation and characterization of cholesterol-independent non-GS NS0 cell lines for production of recombinant antibodies.
AU Hartman T.E.; Sar N.; Genereux K.; Barritt D.S.; He Y.; Burky J.E.; Wesson M.C.; Tso J.Y.; Tsurushita N.; Zhou W.; Sauer P.W.
CS T.E. Hartman, Process Sciences and Engineering, PDL BioPharma, Inc., 34801
Campus Drive, Fremont, CA 94555, United States. taymar.hartman@pdl.com
SO Biotechnology and Bioengineering, (1 Feb 2007) Vol. 96, No. 2, pp. 293-306. .
Refs: 44
ISSN: 0006-3592 E-ISSN: 1097-0290 CODEN: BIBIAU
CY United States
DT Journal; Article
FS 022 Human Genetics
026 Immunology, Serology and Transplantation
LA English
SL English
ED Entered STN: 9 Mar 2007
Last Updated on STN: 9 Mar 2007
AB Presented is an antibody production platform based on the fed-batch culture of recombinant NS0-derived cell lines. NS0 host cells, obtained from the European Collection of Cell Cultures (ECACC, Salisbury, UK, Part No. 85110503), were first adapted to grow in a protein-free, cholesterol-free medium. The resulting host cell line was designated NS0-PFCF (protein-free, cholesterol-free). The five production cell lines presented here were generated using a common protocol consisting of transfection by electroporation and subcloning. The NS0-PFCF host cell line was transfected using a single expression vector containing the Escherichia coli xanthine-guanine phosphoribosyl transferase gene (gpt), and the antibody heavy and light chain genes driven by the ***CMV*** promoter. The five cell lines were chosen after one to three rounds of iterative subcloning, which resulted in a 19-64% increase in antibody productivity when four mother-daughter cell pairs were cultured in a fed-batch bioreactor process. The production cell lines were genetically characterized to determine antibody gene integrity, nucleotide sequences, copy number, and the number of insertion sites in the NS0 cell genome. Genetic characterization data indicate that each of the five production cell lines has a single stably integrated copy of the antibody expression vector, and that the antibody genes are correctly expressed. Stability of antibody production was evaluated for three of the five cell lines by comparing the early stage seed bank with the Working Cell Bank (WCB). Antibody productivity was shown to be stable in two of three cell lines evaluated, while one of the cell lines exhibited a 20% drop in productivity after passaging for approximately 4 weeks. These five NS0-derived production cell lines were successfully cultured to produce antibodies with acceptable product quality attributes in a standardized fed-batch bioreactor process, consistently achieving an average specific productivity of 20-60 pg/cell-day, and a volumetric productivity exceeding 120 mg/L-day (Burky et al., 2006). In contrast to the commonly available NS0 host cell line, which requires serum and cholesterol for growth, and the commonly used expression vector system, which uses a proprietary ***glutamine*** ***synthetase*** selection ***marker*** (GS-NS0), these NS0 cells are cholesterol-independent, grow well in a protein-free medium, use a non-proprietary selection ***marker*** , and do not require gene amplification for productivity improvement. These characteristics are advantageous for use of this NS0 cell line platform for manufacturing therapeutic antibodies. .COPYRGT. 2006 Wiley Periodicals, Inc.

L4 ANSWER 4 OF 9 CAPLUS COPYRIGHT 2007 ACS on STN
AN 2007:95018 CAPLUS <<LOGINID::20070611>>
DN 146:336548
TI Derivation and characterization of cholesterol-independent non-GS NS0 cell lines for production of recombinant antibodies
AU Hartman, Taymar E.; Sar, Nalin; Genereux, Kimberly; Barritt, Diana S.; He, Yimin; Burky, John E.; Wesson, Mark C.; Tso, J. Yun; Tsurushita, Naoya; Zhou, Weichang; Sauer, Paul W.
CS Process Sciences and Engineering, PDL BioPharma, Inc., Fremont, CA, 94555,
USA
SO Biotechnology and Bioengineering (2006), Volume Date 2007, 96(2), 294-306
CODEN: BIBIAU; ISSN: 0006-3592
PB John Wiley & Sons, Inc.
DT Journal
LA English
AB Presented is an antibody prodn. platform based on the fed-batch culture of recombinant NS0-derived cell lines. NS0 host cells, obtained from the European Collection of Cell Cultures (ECACC, Salisbury, UK, Part No. 85110503), were first adapted to grow in a protein-free, cholesterol-free medium. The resulting host cell line was designated NS0-PFCF (protein-free, cholesterol-free). The five prodn. cell lines presented here were generated using a common protocol consisting of transfection by electroporation and subcloning. The NS0-PFCF host cell line was transfected using a single expression vector contg. the Escherichia coli xanthine-guanine phosphoribosyl transferase gene (gpt), and the antibody heavy and light chain genes driven by the ***CMV*** promoter. The five cell lines were chosen after one to three rounds of iterative subcloning, which resulted in a 19-64% increase in antibody productivity when four mother-daughter cell pairs were cultured in a fed-batch bioreactor process. The prodn. cell lines were genetically characterized to det. antibody gene integrity, nucleotide sequences, copy no., and the no. of insertion sites in the NS0 cell genome. Genetic characterization data indicate that each of the five prodn. cell lines has a single stably integrated copy of the antibody expression vector, and that the antibody genes are correctly expressed. Stability of antibody prodn. was evaluated for three of the five cell lines by comparing the early stage seed bank with the Working Cell Bank (WCB). Antibody productivity was shown to be stable in two of three cell lines evaluated, while one of the cell lines exhibited a 20% drop in productivity after passaging for approx. 4 wk. These five NS0-derived prodn. cell lines were successfully cultured to produce antibodies with acceptable product quality attributes in a standardized fed-batch bioreactor process, consistently achieving an av. specific productivity of 20-60 pg/cell-day, and a volumetric productivity exceeding 120 mg/L-day. In contrast to the commonly available NS0 host cell line, which requires serum and cholesterol for growth, and the commonly used expression vector system, which uses a proprietary ***glutamine*** ***synthetase*** selection ***marker*** (GS-NS0), these NS0 cells are cholesterol-independent, grow well in a protein-free medium, use a non-proprietary selection ***marker*** , and do not require gene amplification for productivity improvement. These characteristics are advantageous for use of this NS0 cell line platform for manufg. therapeutic antibodies.
RE.CNT 44 THERE ARE 44 CITED REFERENCES AVAILABLE FOR THIS RECORD
ALL CITATIONS AVAILABLE IN THE RE FORMAT

L4 ANSWER 5 OF 9 CAPLUS COPYRIGHT 2007 ACS on STN
AN 2006:760190 CAPLUS <<LOGINID::20070611>>
DN 145:329461
TI Method for rapidly constructing transgenic mammalian cell line with high-level expression of target gene
IN Huang, Ying; Wang, Yan; Shen, Beifen; Li, Yan
PA Beijing Tianguangshi Biotechnology Co., Ltd., Peop. Rep. China
SO Faming Zhuanli Shenqing Gongkai Shuomingshu, 31pp.
CODEN: CNXXEV
DT Patent
LA Chinese
FAN.CNT 1

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI CN 1693467	A	20051109	CN 2005-10064335	20050414
PRAI CN 2005-10064335		20050414		

AB The invention provides a ***marker*** -integrated vector and a target gene expression vector for high-level expression of the target gene in mammalian cells. The ***marker*** vector contains a dominant selective ***marker*** , a screening ***marker*** , one or more amplification genes, genetic elements for proliferation, and recombination signal sequence (RSS). The expression vector contains a dominant selective ***marker*** , genetic elements, RSS which is identical with that in the ***marker*** vector, a target gene expression cassette, and replication origin of eukaryotic virus. The invention also provides a method for prepg. transgenic mammalian cell lines with stable and high-level expression of the target gene by steps of using the inventive ***marker*** vector to prep. cell lines in which active sites with high transcription level in the genome can be PCR amplified; co-transforming the cell lines with the target gene expression vector and the recombinase expression vector, wherein the recombinase expression vector expresses RSS-specific recombinase, which catalyzes specific recombination at the above two RSS sites; and screening pos. transformants.

L4 ANSWER 6 OF 9 CAPLUS COPYRIGHT 2007 ACS on STN
AN 2004:80885 CAPLUS <<LOGINID::20070611>>
DN 140:140659

TI Vector containing viral cytomegalovirus promoter, mouse IgG2a gene, and ***glutamine*** ***synthetase*** cDNA, and its use in transfecting CHO cells for recombinant protein production
IN Kallmeier, Robert; Gay, Robert
PA Lonza Biologics Plc., UK
SO PCT Int. Appl., 49 pp.
CODEN: PIXXD2
DT Patent
LA English
FAN.CNT 1

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI WO 2004009823	A1	20040129	WO 2003-EP7946	20030721
W: AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NI, NO, NZ, OM, PG, PH, PL, PT, RO, RU, SC, SD, SE, SG, SK, SL, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, YU, ZA, ZM, ZW				
RW: GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZM, ZW, AM, AZ, BY, KG, KZ, MD, RU, TJ, TM, AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HU, IE, IT, LU, MC, NL, PT, RO, SE, SI, SK, TR, BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG				
CA 2489016	A1	20040129	CA 2003-2489016	20030721
AU 2003251434	A1	20040209	AU 2003-251434	20030721
EP 1525320	A1	20050427	EP 2003-765075	20030721
R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT, IE, SI, LT, LV, FI, RO, MK, CY, AL, TR, BG, CZ, EE, HU, SK				
CN 1668749	A	20050914	CN 2003-816914	20030721
JP 2006513696	T	20060427	JP 2004-522537	20030721
IN 2004CN03175	A	20060303	IN 2004-CN3175	20041213
US 2006003405	A1	20060105	US 2005-521768	20050119
PRAI GB 2002-16648	A	20020719		
WO 2003-EP7946	W	20030721		

AB The invention provides a CHO cell transformed with an expression vector comprising a: (a) promoter active in CHO cells and able to express a recombinant protein; (b) portion of the mouse IgG2a gene, which is able to enhance said promoter; and (c) selectable ***marker*** , such as ***glutamine*** ***synthetase*** gene. The invention provides the use of said transfected CHO cell in recombinant prodn. of a protein of interest. The invention relates that said promoter is a strong viral promoter, such as human cytomegalovirus (hCMV) promoter, and that said IgG2a gene lacks its naturally occurring promoter. The invention also relates that mouse ***CMV*** promoter can enhance transfection rate in CHO cells. As way of illustration, green fluorescent protein was expressed in CHO-K1 cells transfected with expression vectors contg. sequences for hCMV promoter and hCMV promoter in presence of IgG2a hot spot sequence. The invention provided the DNA sequences for some of the expression vectors used in this illustration.
RE.CNT 6 THERE ARE 6 CITED REFERENCES AVAILABLE FOR THIS RECORD
ALL CITATIONS AVAILABLE IN THE RE FORMAT

L4 ANSWER 7 OF 9 CAPLUS COPYRIGHT 2007 ACS on STN
AN 2001:50811 CAPLUS <<LOGINID::20070611>>
DN 134:111243
TI Method for selecting high-expressing host cells using dicistronic expression system containing selectable/amplifiable gene within an intron
IN Chisholm, Vanessa; Crowley, Craig W.; Krummen, Lynne A.; Meng, Yu-Ju G.
PA Genentech, Inc., USA
SO PCT Int. Appl., 75 pp.
CODEN: PIXXD2
DT Patent
LA English
FAN.CNT 2

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI WO 2001004306	A1	20010118	WO 2000-US18841	20000711
W: AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CR, CU, CZ, DE, DK, DM, DZ, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW				
RW: GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW, AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG				
CA 2385102	A1	20010118	CA 2000-2385102	20000711
EP 1196566	A1	20020417	EP 2000-945309	20000711
EP 1196566	B1	20060201		
R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT, IE, SI, LT, LV, FI, RO, CY				
JP 2003504059	T	20030204	JP 2001-509510	20000711
AT 317011	T	20060215	AT 2000-945309	20000711
ES 2257303	T3	20060801	ES 2000-945309	20000711
US 2005005310	A1	20050106	US 2003-714000	20031114
US 2007037254	A1	20070215	US 2006-535038	20060925
US 2007054303	A1	20070308	US 2006-535003	20060925
PRAI US 1999-143360P	P	19990712		
WO 2000-US18841	W	20000711		
US 2001-19586	A2	20011220		
US 2003-714000	A1	20031114		

AB Vectors and methods for efficient isolation of recombinant cells expressing high levels of a desired protein are provided. The vectors comprise an amplifiable selectable gene, a fluorescent protein gene, and a gene encoding a desired product in a manner that optimizes transcriptional and translational linkage. The method utilizes eukaryotic host cells harboring a DNA construct comprising a selectable gene (preferably an amplifiable gene) and a product gene provided 3' to the selectable gene. The selectable gene is positioned within an intron defined by a splice donor site and a splice acceptor site and the selectable gene and product gene are under the transcriptional control of a single transcriptional regulatory region. The splice donor site is generally an efficient splice donor site and thereby regulates expression of the product gene using the transcriptional regulatory region. The transfected cells are cultured so as to express the gene encoding the product in a selective medium comprising an amplifying agent for sufficient time to allow amplification to occur, whereupon either the desired product is recovered or cells having multiple copies of the product gene are identified. CHO cells contg. tissue plasminogen activator (tPA) expression vectors according to the invention produced .gtoreq.9-fold higher tPA levels after amplification than did CHO cells contg. conventional vectors. The vector was a pRK deriv. This vector contains a cytomegalovirus immediate early promoter and an intron having a splice donor site derived from the cytomegalovirus immediate early gene and a splice acceptor site from an IgG heavy chain variable region gene. The DHFR gene was inserted into this intron and the tPA gene was inserted downstream of the splice acceptor site.
RE.CNT 7 THERE ARE 7 CITED REFERENCES AVAILABLE FOR THIS RECORD
ALL CITATIONS AVAILABLE IN THE RE FORMAT

L4 ANSWER 8 OF 9 CAPLUS COPYRIGHT 2007 ACS on STN
AN 1999:779157 CAPLUS <<LOGINID::20070611>>
DN 132:19632
TI Method for integrating genes at specific sites in mammalian cells via homologous recombination and vectors for accomplishing the same
IN Reff, Mitchell R.; Barnett, Richard Spence; McLachlan, Karen Retta
PA Idec Pharmaceuticals Corporation, USA
SO U.S., 43 pp., Cont.-in-part of U.S. 5,830,698.
CODEN: USXXAM
DT Patent
LA English
FAN.CNT 2

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI US 5998144	A	19991207	US 1998-23715	19980213
US 5830698	A	19981103	US 1997-819866	19970314
CA 2283740	A1	19980924	CA 1998-2283740	19980309
CA 2283740	C	20060627		
WO 9841645	A1	19980924	WO 1998-US3935	19980309
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RW: GH, GM, KE, LS, MW, SD, SZ, UG, ZW, AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG				
AU 9864435	A	19981012	AU 1998-64435	19980309
AU 737155	B2	20010809		
EP 981637	A1	20000301	EP 1998-910109	19980309
EP 981637	B1	20050525		
R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT, IE, FI				
BR 9808584	A	20000523	BR 1998-8584	19980309
HU 200002320	A2	20001128	HU 2000-2320	19980309
JP 2001516221	T	20010925	JP 1998-540539	19980309
CZ 293355	B6	20040414	CZ 1999-3162	19980309
AT 296356	T	20050615	AT 1998-910109	19980309
PT 981637	T	20050930	PT 1998-910109	19980309
RO 120148	B1	20050930	RO 1999-972	19980309
ES 2242997	T3	20051116	ES 1998-910109	19980309
EP 1605054	A1	20051214	EP 2005-75757	19980309
R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT, IE, FI				
EP 1605055	A1	20051214	EP 2005-76212	19980309
R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT, IE, FI				
PL 191251	B1	20060428	PL 1998-335695	19980309
IN 189732	A1	20030419	IN 1998-DE616	19980310
ZA 9802152	A	19981009	ZA 1998-2152	19980313
TW 232239	B	20050511	TW 1998-87103778	19980313
US 6413777	B1	20020702	US 1999-343485	19990630
NO 9904397	A	19991109	NO 1999-4397	19990910
MX 9908363	A	20000630	MX 1999-8363	19990910
US 2002192820	A1	20021219	US 2002-109853	20020401
US 6841383	B2	20050111		
US 2004166528	A1	20040826	US 2004-817950	20040406
IN 2004DE02321	A	20060922	IN 2004-DE2321	20041119
PRAI US 1997-819866	A2	19970314		
US 1998-23715	A	19980213		
EP 1998-910109	A3	19980309		
WO 1998-US3935	W	19980309		
IN 1998-DE616	A3	19980310		
US 1999-343485	A1	19990630		

US 2002-109853 A1 20020401
AB A method for achieving site specific integration of a desired DNA at a target site in a mammalian cell via homologous recombination is described. This method provides for the reproducible selection of cell lines wherein a desired DNA is integrated at a predetd. transcriptionally active site previously marked with a ***marker*** plasmid (Desmond). This unique site may be bacterial DNA, a viral DNA or synthetic DNA. This Desmond ***marker*** plasmid contains the Salmonella HisD gene, the Neomycin phosphotransferase exon 3, the murine dihydrofolate reductase, cytomegalovirus and SV40 enhancers, splice acceptor site, mouse beta globin major promoter, bovine growth hormone polyadenylation site, SV40 early and late polyadenylation sites. The selectable ***marker*** proteins may include neomycin phosphotransferase, histidinol dehydrogenase, dihydrofolate reductase, hygromycin phosphotransferase, HSV thymidine kinase, adenosine deaminase, ***glutamine*** ***synthetase*** , and hypoxanthine-guanine phosphoribosyl transferase. Marked CHO cells were produced and characterized. Other cells that may be marked include myeloma cells, baby hamster kidney cells, COS cells, NSO cells, HeLa cells and NIH 3T3 cells. The method is particularly suitable for the prodn. of mammalian cell lines which secrete mammalian proteins at high levels, in particular Igs. Novel targeting vectors (Molly) and vector combinations for use in the subject cloning method are also provided. This Molly vector contains dihydrofolatereductase, N1+Neomycin phosphotransferase exon1, N2+Neomycin phosphotransferase exon 2, anti-CD20 light chain leader+variable, human kappa const., anti-CD20 heavy chain leader+variable, human gamma 1 const., Salmonella histidinol dehydrogenase, ***CMV*** and SV40 enhancers, SV40 origin, splice donor/acceptor, ***CMV*** promoter/enhancer, HSV TK promoter and poloma enhancer, mouse beta globin major promoter, SV40 late polyadenylation, bovine growth hormone polyadenylation. Expression of an Anti-CD20 and Anti-human CD23 antibody and immunoadhesin in Desmond marked CHO cells was achieved.
RE.CNT 1 THERE ARE 1 CITED REFERENCES AVAILABLE FOR THIS RECORD
ALL CITATIONS AVAILABLE IN THE RE FORMAT

L4 ANSWER 9 OF 9 CAPLUS COPYRIGHT 2007 ACS on STN
AN 1998:228730 CAPLUS <<LOGINID::20070611>>
DN 129:63681
TI High-level expression of HBsAg in CHO cells using ***glutamine*** ***synthetase*** gene as an amplifiable selectable ***marker***
AU Liu, Wenjun; Yang, Furong; Ruan, Li; Ren, Guifang; Zhu, Jiming
CS Institute of Virology, Chinese Academy of Preventive Medicine, Beijing, 100052, Peop. Rep. China
SO Bingdu Xuebao (1997), 13(2), 103-109
CODEN: BIXUEA; ISSN: 1000-8721
PB Bingdu Xuebao Bianjibu
DT Journal
LA Chinese
AB ***Glutamine*** ***synthetase*** gene (GS) was introduced into CHO cells. The transfectants were selected by growth in a glutamine-free medium. Vector amplification was subsequently selected using specific inhibitor of GS, methionine sulfoximine (MSX). Combining with the ***CMV*** promoter, HBV S gene were expressed in CHO cells. After two rounds of selection for vector amplification, the expression level of G4 cell line was >1:256 (RPHA), which was twice of that of the B43 cell line obtained using dhfr gene amplification.

=>

---Logging off of STN---

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Executing the logoff script...

=> LOG Y

COST IN U.S. DOLLARS	ENTRY	SINCE FILE SESSION	TOTAL
FULL ESTIMATED COST		36.32	37.37

DISCOUNT AMOUNTS (FOR QUALIFYING ACCOUNTS)	ENTRY	SESSION	SINCE FILE TOTAL
CA SUBSCRIBER PRICE		-5.46	-5.46

STN INTERNATIONAL LOGOFF AT 16:44:37 ON 11 JUN 2007